

Synthesis of 4-Thiopseudoisocytidine and 4-Thiopseudouridine as Components of Triplex-forming Oligonucleotides

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In this paper, we report convenient methods for the synthesis of 4-thiopseudoisocytidine ($s^4\psi$ iC) and 4-thiopseudouridine ($s^4\psi$ U). ^1H NMR spectral analysis of these modified nucleosides showed that both $s^4\psi$ U and $s^4\psi$ iC prefer C3'-*endo* ribose puckering. These conformational properties are favorable for the stabilization of triplex formation.

Using the antigene strategy, a large number of modified nucleosides have been synthesized to enhance the thermal stability of DNA triplexes formed by hybridization of the third DNA strands with DNA duplexes.^{1–5} These studies showed that the use of homopyrimidine–oligodeoxynucleotides containing cytosine or 5-methylcytosine bases as triplex-forming oligodeoxynucleotides (TFOs) under weakly acidic conditions resulted in significant stabilization of the resulting parallel triplex structures. This was due to the formation of protonated cytosine or 5-methylcytosine bases that could bind to guanine bases at the Hoogsteen base-pairing site.^{6–9} However, those acidic conditions limit the sequences of TFOs; therefore, antigene therapy using this strategy is not generally applicable. To overcome this limitation, several modified nucleosides have been developed to mimic the structure of the 3-N-protonated cytosine base.^{10–15} 2'-O-Methylpseudoisocytidine (ψ iCm) is known to form a triplet base pair with a G–C base pair under neutral conditions. However, TFOs containing ψ iCm could not stabilize the triplex structure sufficiently at neutral pH.^{10,11}

On the other hand, we have recently reported that TFOs containing 2'-O-methyl-2-thiouridine (s^2 Um) or 2-thiothymidine (s^2 T) formed quite stable parallel triplex structures.¹⁶ Enhancement of the thermal stability of these parallel triplexes can be explained by means of the strong stacking interaction of the 2-thiocarbonyl group with the 5'-upstream or 3'-downstream bases. In particular, it was found that a consecutive alignment of s^2 Um or s^2 T in TFOs resulted in a more effective increase in the binding ability toward DNA duplexes.¹⁶

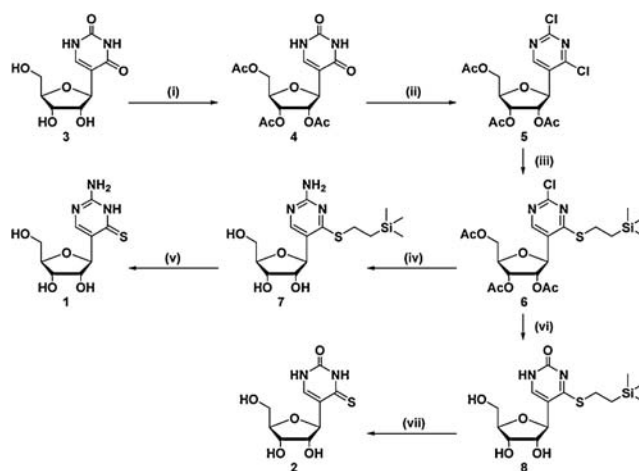
It was expected that a consecutive pile of 4-thiopseudoisocytidine (**1**: $s^4\psi$ iC) in combination with s^2 Um or s^2 T might cause an increase in the thermal stability of the parallel triplex structures. In this paper, we report convenient methods for the synthesis of **1** and 4-thiopseudouridine (**2**: $s^4\psi$ U), which can be derived from a synthetic intermediate of the former. Chemical structures of these modified nucleosides were shown in Figure 1.



Figure 1. Chemical structures of 4-thiopseudoisocytidine and 4-thiopseudouridine.

In the synthesis of 4-thiouridine (s^4 U), it was reported that the thiolation of the pyrimidine ring at position 4 could be achieved by the reaction of 4-(2,4,6-triisopropylbenzenesulfonyl)pyrimidinone nucleoside derivatives with 3-sulfanypropionitrile.^{17–19} In addition, many reactions with pyrimidine rings substituted with leaving groups at position 4 were reported. Therefore, such types of substitution reactions might also produce 4-substituted ψ derivatives. Townsend et al. reported 2,4-dichloro-5-(2,3,5-tri-O-acetyl- β -D-ribofuranosyl)pyrimidine as a ψ derivative that has chloro groups at positions 2 and 4 on the pyrimidine ring.²⁰ Considering the reactivity of compound, we expected that the substitution reaction might occur predominantly at position 4.²¹ According to Townsend's procedure (Scheme 1), pseudouridine **3** was converted to 2',3',5'-tri-O-acetylpsudouridine (**4**) in 92% yield. Compound **4** was further treated with excess POCl_3 to give 2,4-dichloropsudouridine derivative **5** in 91% yield.

After that, as expected, the reaction of compound **5** with 2-(trimethylsilyl)ethanethiol in *N,N*-dimethylacetamide in the presence of triethylamine formed only the 4-thiolated compound **6** in a high yield of 84%. The structure of this product was determined from the correlation between the ^1H signal of the 2-(trimethylsilyl)ethyl group and the ^{13}C signal of 4C on the pyrimidine ring, obtained by HMBC spectrum analysis. The chloro group of compound **6** was converted to an amino group by the reaction with concd NH_3 to form compound **7** in 44% yield. Treatment of **7** with Bu_4NF formed $s^4\psi$ iC (**1**) in 62% yield.²²

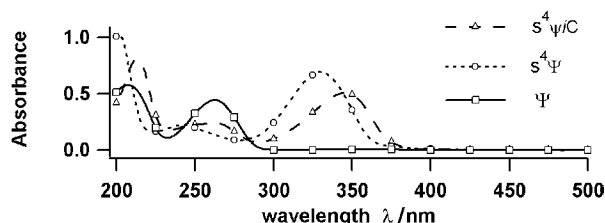


Scheme 1. Reagents and conditions: (i) Ac_2O (10 equiv), pyridine, rt; (ii) *N,N*-diethylamine hydrochloride (1.0 equiv), POCl_3 (20 equiv), reflux; (iii) 2-(trimethylsilyl)ethanethiol (1.2 equiv), triethylamine (1.2 equiv), DMA, rt; (iv) concd NH_3 , dioxane, 100 °C; (v) TBAF (3.0 equiv), THF, 50 °C; (vi) $\text{LiOH}\cdot\text{H}_2\text{O}$ (5.0 equiv), DMA, 60 °C; and (vii) TBAF (1.5 equiv), THF, 60 °C.

Table 1. Conformational analysis of $s^4\Psi$ and $s^4\psi iC$ in D_2O

	Ψ	$s^4\Psi$	$s^4\psi iC$
%N (C3'-endo) ^a	50%	78%	66%
$J_{1'H2'H}$	5.4 Hz	2.2 Hz	3.9 Hz
$J_{3'H4'H}$	5.4 Hz	7.8 Hz	7.1 Hz

^a%N values of nucleosides were determined by following equation: %N (C3'-endo) = $J_{3'H4'H}/(J_{1'H2'H} + J_{3'H4'H}) \times 100$.

**Figure 2.** UV spectra of $s^4\psi iC$ and $s^4\Psi$ in H_2O .

On the other hand, hydrolysis of compound **6** with LiOH afforded 4-(2-trimethylsilyl)ethyl-4-thiopseudouridine (**8**) in 36% yield. The low yields of the above two reactions of compound **6** forming compounds **7** and **8** were due to side reactions of position 4, since it is known that pyrimidine derivatives having alkylthio or sulfanyl groups at position 4 or 2 react easily with nucleophilic reagents.^{23–26} The TBAF-mediated deprotection of compound **8** formed $s^4\Psi$ in 67% yield.²⁷

To clarify the sugar conformations of $s^4\Psi$ and $s^4\psi iC$, 1H NMR spectral analysis was performed. As shown in Table 1, it was found that $s^4\Psi$ and $s^4\psi iC$ showed C3'-endo ribose puckering forms (%N; $s^4\Psi$: 78%; $s^4\psi iC$: 66%) more predominantly than Ψ . It was reported that s^2U derivatives prefer C3'-endo ribose puckering.^{28–30} This conformational predominance is known to be caused by steric repulsion between the 2-thiocarbonyl group of s^2U and the 2'-hydroxy group.²⁸ The C3'-endo predominance observed could be explained by the same type of steric repulsion.

It is known that s^4U exhibited a unique UV absorption spectrum with maximum absorbance at 330 nm.³¹ As shown in Figure 2, the UV absorption maxima of 4-thiopseudo-nucleosides $s^4\psi iC$ and $s^4\Psi$ were shifted markedly from that of Ψ (260 nm) to 345 and 331 nm, respectively. These spectral changes were very similar to those from U to s^4U . Since the structure of s^4U resembles that of $s^4\Psi$, these UV spectral changes also supported the view that the thiolation occurred at position 4 of the pyrimidine ring.

In conclusion, we synthesized $s^4\psi iC$ and $s^4\Psi$ successfully. The 1H NMR studies of these modified nucleosides showed that both $s^4\psi iC$ and $s^4\Psi$ prefer C3'-endo ribose puckering. These conformational properties are favorable for the stabilization of both RNA-duplex and parallel triplex formation. Synthesis of oligonucleotides containing $s^4\psi iC$ and study of their duplex- and triplex-forming abilities are now in progress.

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References and Notes

- C. Hélène, J.-J. Toulme, *Biochim. Biophys. Acta* **1990**, 1049, 99.
- D. Praseuth, A. L. Guieysse, C. Hélène, *Biochim. Biophys. Acta* **1999**, 1489, 181.
- K. R. Fox, *Curr. Med. Chem.* **2000**, 7, 17.
- S. Buchini, C. J. Leumann, *Curr. Opin. Chem. Biol.* **2003**, 7, 717.
- R. V. Guntaka, B. R. Varma, K. T. Weber, *Int. J. Biochem. Cell Biol.* **2003**, 35, 22.
- C. Des los Santos, M. Rosen, D. Patel, *Biochemistry* **1989**, 28, 7282.
- P. Rajagopal, J. Feigon, *Biochemistry* **1989**, 28, 7859.
- J. L. Asensio, A. N. Lane, J. Dhesi, S. Bergqvist, T. Brown, *J. Mol. Biol.* **1998**, 275, 811.
- J. Robles, A. Grandas, E. Pedroso, F. J. Luque, R. Eritja, M. Orozco, *Curr. Org. Chem.* **2002**, 6, 1333.
- A. Ono, P. O. P. Ts'o, L. S. Kan, *J. Am. Chem. Soc.* **1991**, 113, 4032.
- A. Ono, P. O. P. Ts'o, L. S. Kan, *J. Org. Chem.* **1992**, 57, 3225.
- G. Xiang, W. Soussou, L. W. McLaughlin, *J. Am. Chem. Soc.* **1994**, 116, 11155.
- R. Berressem, J. W. Engels, *Nucleic Acids Res.* **1995**, 23, 3465.
- U. von Krosigk, S. A. Benner, *J. Am. Chem. Soc.* **1995**, 117, 5361.
- G. Xiang, R. Bogacki, L. W. McLaughlin, *Nucleic Acids Res.* **1996**, 24, 1963.
- I. Okamoto, K. Seio, M. Sekine, *Bioorg. Med. Chem. Lett.* **2006**, 16, 3334.
- R. S. Coleman, J. M. Siedlecki, *Tetrahedron Lett.* **1991**, 32, 3033.
- C. J. Adams, J. B. Murray, J. R. P. Arnold, P. G. Stockley, *Tetrahedron Lett.* **1994**, 35, 765.
- R. S. Coleman, E. A. Kesicki, *J. Am. Chem. Soc.* **1994**, 116, 11636.
- 2,4-Dichloro-5-(β -D-ribofuranosyl)pyrimidines and substituted derivatives: L. B. Townsend, D. S. Wise, R. A. Earl, G. Belton, U.S. Pat. Appl. 4092472, **1989**.
- R. G. Shepherd, J. L. Fedrick, *Adv. Heterocycl. Chem.* **1965**, 4, 145.
- Physical properties of 4-thiopseudoisocytidine (**1**) are shown in Supporting Information. Supporting Information is available electronically on the CSJ-Journal Web site, <http://www.csj.jp/journals/chem-lett/index.html>.
- J. J. Fox, D. Van Praag, I. Wempen, I. L. Doerr, L. Cheong, J. E. Knoll, M. L. Eidinoff, A. Bendich, G. B. Brown, *J. Am. Chem. Soc.* **1959**, 81, 178.
- C.-H. Niu, *Anal. Biochem.* **1984**, 139, 404.
- T. Ueda, J. J. Fox, *J. Med. Chem.* **1963**, 6, 697.
- H. C. Van der Plas, B. Zuurdeeg, H. W. van Meeteren, *Recl. Trav. Chim. Pays-Bas* **1969**, 88, 1156.
- Physical properties of 4-thiopseudouridine (**2**) are shown in Supporting Information. Supporting Information is available electronically on the CSJ-Journal Web site, <http://www.csj.jp/journals/chem-lett/index.html>.
- Y. Yamamoto, S. Yokoyama, T. Miyazawa, K. Watanabe, S. Higuchi, *FEBS Lett.* **1983**, 157, 95.
- H. Sierzputowska-Gracz, E. Sochacka, A. Malkiewicz, K. Kuo, C. W. Gehrke, P. F. Agris, *J. Am. Chem. Soc.* **1987**, 109, 7171.
- P. F. Agris, H. Sierzputowska-Gracz, W. Smith, A. Malkiewicz, E. Sochacka, B. Nawrot, *J. Am. Chem. Soc.* **1992**, 114, 2652.
- W. Saenger, *Principles of Nucleic Acid Structure*, Springer-Verlag, New York, **1987**.